ORIGINAL RESEARCH article

Sorghum bicolor-based supplement reduces oxidative stress and pro-inflammatory cytokines to mitigate rotenone-induced Parkinsonian-like motor dysfunctions in rats

Paul A. Adeleke ¹ 🖾 (b), Olajide S. Annafi ² 🖾 (b), Abayomi M. Ajayi ¹ 🖾 (b) Benneth Ben-Azu ³ 🖾 (b), Olajuwon Okubena ⁴ 🖾 (b), and Solomon Umukoro ^{1*} 🖾 (b)

¹ Department of Pharmacology and Therapeutics, College of Medicine, University of Ibadan, Sango-Ojo Road, Ibadan, Oyo State, ² Department of Pharmacology and Therapeutics, College of Medicine, Osun State University, Osun, ³ DELSU Joint Canada-Israel Neuroscience and Biopsychiatry Laboratory, Department of Pharmacology, Faculty of Basic Medical Sciences, College of Health Sciences, Delta State University, Abraka, Delta State, ⁴ Health Forever Products Inc., Lagos, Nigeria * Author to whom correspondence should be addressed

Received: 22-07-2024, Revised: 12-08-2024, Accepted: 15-08-2024, Published: 30-09-2024

Copyright[©] 2024. This open-access article is distributed under the *Creative Commons Attribution License*, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

HOW TO CITE THIS

Adeleke et al. (2024) *Sorghum bicolor*-based supplement reduces oxidative stress and pro-inflammatory cytokines to mitigate rotenone-induced Parkinsonian-like motor dysfunctions in rats. Mediterr J Pharm Pharm Sci. 4 (3): 15-26. [Article number: 165]. https://doi.org/10.5281/zenodo.13309953

Keywords: Pro-inflammatory cytokines, motor deficits, oxidative stress, rotenone

Abstract: Parkinson's disease is a common movement disorder associated primarily with oxidative stressmediated degeneration of dopaminergic neurons. Earlier studies showed that *Sorghum bicolor*-based supplement (SbS) exhibited antioxidant and neuroprotective activities and might likely rescue the death of dopaminergic neurons in Parkinson's disease. This study examined the effect of SbS on rotenone-induced Parkinsonian-like motor deficits in rats and the involvement of oxidative stress and pro-inflammatory cytokines. Rats were divided into six groups and treated orally with sunflower oil (vehicle-control), rotenone (2.5 mg/kg) alone or in combination with each dose of SbS (50, 100, and 200 mg/kg) and levodopa-carbidopa (10 mg/kg) on an alternate day for 28 days. The changes in motor functions were evaluated on day 28 and the brain concentrations of oxidative stress biomarkers and pro-inflammatory cytokines (tumor necrosis factoralpha and interleukin-6) were determined. Rotenone caused motor deficits by impaired locomotor activity in the open field test and induced catalepsy in the bar test, which were attenuated by SbS. Rats pretreated with SbS had reduced brain levels of malondialdehyde, nitrite, and pro-inflammatory cytokines compared to rotenone controls. SbS mitigated rotenone-induced depletion of reduced glutathione and antioxidant enzymes in the rat brain. The results suggest that SbS ameliorated rotenone-induced Parkinsonian-like motor dysfunctions by reducing neuronal oxidative stress and pro-inflammatory cytokines in rats.

Introduction

Parkinson's Disease (PD) has been described as one of the most common neurodegenerative disorders after Alzheimer's disease (AD), which is characterized by the loss of dopamine neurons in the substantia nigra and the presence of Lewy bodies in surviving dopaminergic neurons [1, 2]. The motor symptoms of PD include tremors at rest, muscle rigidity, slowness of movement and postural instability [3, 4]. As PD progresses, walking becomes a difficult task for the patient due to postural instability and the tendency to fall. Thus, in the long term, the quality of life of the patient is grossly impaired and he becomes dependent on others for

Mediterranean Journal of Pharmacy & Pharmaceutical Sciences www.medjpps.com

daily activities [5]. Oxidative stress and inflammation are strongly implicated in the pathophysiology of PD and the progression of the illness [5, 6]. For example, oxidative stress has been identified as the prime factor in initiating cellular injury and death of dopaminergic neurons in PD [7, 8]. Studies have reported elevated levels of oxidative stress in postmortem brain tissue of patients with PD, including increased cholesterol lipid hydroperoxides in the substantia nigra [9-12]. Increased cytoplasmic 8-hydroxy-guanosine immunoreactivity and oxidative damage to several brain mitochondrial complex I proteins accompanied by decreased glutathione in neurons of the substantia nigra have been reported [12, 13]. The findings of reduced antioxidant molecules and high concentrations of oxidized lipids and protein in the brains of individuals with PD further confirmed the key role of oxidative stress in the pathology of the illness [6, 12, 13]. Oxidative stress-mediated injury to neuronal cells evokes the release of pro-inflammatory cytokines, thereby linking oxidative stress and inflammation as co-conspirators in the pathogenesis of PD [2, 5, 14, 15]. The key role of oxidative stress and neuroinflammation in the pathophysiology of PD has also been reinforced by the findings that rotenone, a neurotoxin, is widely used to replicate the behavioral and biochemical changes including synucleinopathies akin to PD pathology. Specifically, rotenone is known to cause Parkinsonian-like symptoms through the formation of free radicals that lead to impairment of mitochondrial electron transport machinery [3]. Thus, it has been proposed that since a multifactorial cascade of pathogenic events triggered by oxidative stress causes cell death in PD, the neuroprotective strategy might be a better option for treating PD [14, 16]. Accordingly, oxidative and neuroinflammation have become important advancements and promising targets, with antioxidant and anti-inflammatory agents as promising, reliable therapeutic interventions [17, 18].

Sorghum bicolor-based supplement (SbS) is a supplement obtained from the leaf sheath of *Sorghum bicolor* (Poaceae) that has been shown to exhibit potent antioxidant, neuroprotective and anti-inflammatory activities [19-21]. SbS improved neurological deficits in rats with ischemic stroke by reducing of brain contents of proinflammatory cytokines and expression of NF-kB immunopositive cells [22, 23). In a recent study using *Drosophila melanogaster*, SbS extended the lifespan and improved the motor function of the flies through augmentation of the antioxidant status [23]. In this study, we examined the effect of SbS on rotenone-induced Parkinsonian-like motor deficits in rats and the probable involvement of oxidative stress, and proinflammatory cytokines.

Materials and methods

Chemicals: Rotenone, thiobarbituric acid (TBA) were purchased from Sigma Aldrich (Germany), 5,5'-dithiobis-2-nitrobenzoic acid (Ellman's reagent) and acetylthiocholine iodide from Sigma Aldrich (USA). ELISA kits for rats' tumour necrosis factor-alpha and interleukin-6 from Biolegend (USA). The levodopa-cabidopa (LD-CD) was a product of Merck & Co, Inc (USA).

Experimental animals: Male Albino Wistar rats (weight: 170 g - 200 g, age: 10-13 weeks) used in this study were obtained from the Central Animal House, University of Ibadan, Nigeria. Rats were acclimatized in the Department of Pharmacology and Therapeutics Animal Holding Facility for two weeks before the study. They were housed in cages with free access to a standard rodent pellet diet (Vital Feeds, Jos, Nigeria) and water *ad libitum*. The procedures were by the National Institute of Health (NIH Publication No 8523, revised 1981) guidelines for the Care and Use of Laboratory Animals. Approval was obtained from the University of Ibadan Animal Care and Use Research Ethics Committee (UI-ACUREC/20/0055).

Preparation of SbS and rotenone: SbS was obtained from Health Forever Products Ltd, Lagos, Nigeria and was prepared as described [20]. Briefly, on the experimental day, 500 mg of SbS was dissolved in 25 mL of water to obtain 20 mg/mL. The doses of 50, 100 and 200 mg/kg were selected based on an earlier study [22]. Rotenone was prepared according to a previous study [24]. Thus, rotenone was initially dissolved in dimethyl

sulfoxide (DMSO) and the solution was diluted in sunflower vegetable oil to obtain 0.25 mg per mL. The intraperitoneal dose of 2.5 mg/kg of rotenone was chosen based on an earlier study [24].

Effect of SbS on rotenone-induced Parkinsonian-like symptoms in rats: The effect of SbS on rotenone-induced Parkinsonian-like symptoms was studied in rats according to a previous study [24]. Thus, rats were randomly divided into six groups (n=7). Rats in group 1, which served as vehicle control, received sunflower oil (10 mL/kg, p.o.), group 2 had sunflower oil (rotenone control), groups 3-5 were pre-treated SbS (50, 100 and 200 mg/kg, p.o.) while the last group received LD-CD (10 mg/kg, p.o.), daily for 28 days. 30 min after each pretreatment, rats in groups 2-6 received rotenone (2.5 mg/kg, i.p.) on alternate days for 28 days. Afterward, the test for locomotion and catalepsy using an open field and bars, respectively, were carried out to evaluate the motor functions of the rats on day 28.

Procedures for evaluation of motor functions

Test for spontaneous motor activity (SMA): The SMA was assessed using an open-field test. The open-field apparatus consists of plywood (72x72x36 cm) with the floor divided into sixteen squares, 18x18 cm [25]. The rats were placed individually at the center of the chamber, and the number of lines crossed, and duration of ambulation were recorded for five minutes using a video camera [25].

Test for catalepsy: The effect of SbS on rotenone-induced catalepsy was investigated according to the modified method [26]. The test was done by placing the forelimbs of each rat on a horizontal plane wood surface (H: 6, W: 4, and L: 16 cm) and the duration of akinesia (the period the rat remained in one position, before initiating any active movement) was recorded for five minutes.

Preparation of brain tissues for biochemical assays: Immediately after the behavioral studies, the rats were euthanized using diethyl ether and each of the isolated brains was weighed and rinsed with 10.0% w/v sodium phosphate (0.1 M; pH 7.4). Each brain tissue was homogenized and centrifuged, and the supernatant was collected for the biochemical parameters [22].

Estimation of oxidative stress biomarker and nitrite content: The malondialdehyde (MDA) contents in the brain regions were estimated using the procedure of thiobarbituric reacting substance (TBARS) [27]. The striatal MDA content was calculated using a molar extinction coefficient of 1.56×10.0^5 per M per cm and expressed as µmol MDA per g tissue. The determination of nitrite content is estimated as described [28]. The absorbance was read at 540 nm and the nitrite in the striatum was estimated from the standard curve of sodium nitrite (0.0 M-100 M).

Estimation of antioxidant biomarkers: The brain-reduced glutathione (GSH) was assayed as described previously and expressed as μ M GSH per g tissue [29]. The method described by Goth [30] was utilized for the determination of catalase activity and expressed as μ moles of hydrogen peroxide decomposed (Unit/mg protein). A method of Misra and Fridovich [31] was used for the estimation of superoxide dismutase (SOD) activity (Unit/mg protein). Protein content was estimated using the procedure of Lowry et al. [32].

Determination of tumor necrosis factor-alpha ($TNF-\alpha$) and interleukin-6 (IL-6) contents: The brain concentrations of TNF- α and IL-6 were determined using ELISA kits according to the manufacturer's instructions guide. The concentrations were determined from their standard curves respectively and expressed as pg/mg protein.

Statistical analysis: After normality and homogeneity data checks, data were expressed as mean±S.E.M, and analyzed using Graph Pad Prism software version 9.00 (San Diego, CA, USA). The analysis of data was done using one-way ANOVA, followed by Bonferroni post-hoc test. A p-value less than 0.05 was considered significant.

Results

SBS reduces rotenone-induced locomotor deficits: The effects of SbS on rotenone-induced Parkinsonian-like motor features in rats are shown in **Figures 1** and **2**. **Figure 1** showed that 2.5 mg/kg rotenone reduced the distance traveled $[F_{(5, 18)}=46.84, p<0.001]$ and the speed $[F_{(5, 18)}=34.69, p<0.001]$ in navigating the open field chamber relative to controls. As shown in **Figure 2**, 2.5 mg/kg rotenone increased the freezing episodes (periods in which the rat stops movement) $[F_{(5, 18)}=26.09, p<0.001]$ and the freezing time (period of absence of movement) $[F_{(5, 18)}=74.12, p<0.001]$, and the number of lines crossed when compared with controls $[F_{(5, 18)}=98.72, p<0.001]$. However, an oral administration of 100 mg/kg and 200 mg/kg SbS or 10.0 mg/kg LD-CD significantly improved rat motor functions when compared with rotenone (**Figures 1** and **2**).





Bars are mean \pm S.E.M. # > 0.05 versus vehicle, * > 0.05 versus rotenone (ANOVA followed by Bonferroni test) VEH=vehicle, SbS=Sorghum bicolor supplement, LD-CD=levodopa-carbidopa

Figure 2: Sorghum bicolor supplement mitigated rotenone-induced freezing behaviors and impaired rat locomotion



Bars are mean±*S.E.M.* [#]*p* <0.05 versus vehicle, ^{*}*p*<0.05 versus rotenone (ANOVA followed by Bonferroni test) *VEH*=vehicle, *SbS*=Sorghum bicolor supplement, LD-CD=levodopa-carbidopa

SBS attenuates rotenone-induced catalepsy: In **Figure 3**, intraperitoneal injection of 2.5 mg/kg rotenone caused catalepsy, evidenced by a significant increase in latency to initiation of movement relative to control. An oral dose of 200 mg/kg SbS or 10.0 mg/kg LD-CD highly significantly prevented catalepsy relative to the rotenone group [$F_{(5, 36)}$ =19.36, p<0.001]. In **Figure 3**, lower doses of 50.0 mg/kg SbS to 100 mg/kg SbS could not alter rotenone-induced cataleptic condition.

SBS decreases brain malondialdehyde (MDA): Intraperitoneal injection of 2.5 mg/kg rotenone significantly elevated MDA concentrations of the hippocampus (HIP), prefrontal cortex (PFC) and striatum (STR) regions of rats relative to the control (**Figure 4**). As presented in **Figure 4**, 100 mg/kg and 200 mg/kg of SbS or 10.0 mg/kg LD-CD given orally significantly suppressed MDA concentrations in these brain regions of rotenone-treated rats. [HIP: $F_{(5, 20)}$ =15.36, p<0.001; PFC: $F_{(5, 20)}$ =9.688 p<0.001; STR: $F_{(5, 20)}$ =7.446, p<0.001].

SBS increases glutathione (GSH) in rotenone-treated rats: As presented in **Figure 5**, 2.5 mg/kg rotenone significantly decreased GSH levels in the hippocampus and striatum of rats relative to the control. Though, 100 mg/kg and 200 mg/kg SbS or 10.0 mg/kg LD-CD significantly mitigated rotenone-induced GSH depletion (**Figure 5**) [HIP: $F_{(5, 20)}$ =14.79, p<0.001; STR: $F_{(5, 21)}$ =14.37, p<0.001].





Bars are mean \pm S.E.M. # p<0.05 versus vehicle, * p<0.05 versus rotenone (One-way ANOVA followed by Bonferroni test) VEH=vehicle, SbS=Sorghum bicolor supplement, LD-CD=levodopa-carbidopa





Bars are mean \pm S.E.M. #p<0.05 versus vehicle. *p<0.05 versus rotenone (ANOVA followed by Bonferroni test) VEH=Vehicle, SbS=Sorghum bicolor supplement, LD-CD=Levodopa-carbidopa





GSH



Bars are mean±S.*E.M.* [#]*p*<0.05 versus vehicle. ^{*}*p*<0.05 relative to rotenone (ANOVA followed by Bonferroni test) VEH=vehicle, SbS=Sorghum bicolor supplement, LD-CD=levodopa-carbidopa

SBS boosts neuronal antioxidant activity in rotenone-treated rats: The effects of SbS on antioxidant enzymes (superoxide dismutase-SOD and catalase) in specific brain regions of rats injected with rotenone are presented in **Figures 6** and **7**. Intraperitoneal dose of rotenone (2.5 mg/kg) caused significant suppression of SOD and catalase activities in specific brain regions of rats when compared with control. Though, 100 mg/kg and 200 mg/kg SbS or 10.0 mg/kg LD-CD significantly increased SOD and catalase activity (**Figure 6** and **7**) [$F_{(5, 59)}$ =113.9, p<0.001].





Bars are mean±S.E.M. [#]p<0.05 versus vehicle. ^{*}p<0.05 versus rotenone (ANOVA followed by Bonferroni test) VEH=Vehicle, SbS=Sorghum bicolor Supplement, LD-CD=Levodopa-carbidopa







Bars are mean \pm S.E.M. # p < 0.05 versus vehicle. * p < 0.05 versus rotenone (ANOVA followed by Bonferroni test) VEH=vehicle, SbS=Sorghum bicolor supplement, LD-CD=levodopa-Carbidopa

SBS reduces nitrite contents of rotenone-treated rats: In **Figure 8**, 2.5 mg/kg rotenone significantly increased nitrite concentrations in the hippocampus, striatum and prefrontal cortex of rats compared with the controls. However, 100 mg/kg and 200 mg/kg SbS or 10.0 mg/kg LD-CD significantly reduced nitrite contents in these brain regions of rats relative to rotenone (**Figure 8**) [HIP: $F_{(5, 20)}=12.87$, p<0.001; PFC: $F_{(5, 20)}=17.29$, p<0.001; STR: $F_{(5, 21)}=53.08$, p<0.001].





Nitrite

Bars are mean \pm S.E.M. # p < 0.05 versus vehicle. * p < 0.05 versus rotenone (ANOVA followed by Bonferroni test) VEH=vehicle, SbS=Sorghum bicolor supplement, LD-CD=levodopa-carbidopa



SBS reduces tumornecrosis factor- α and interleukin-6 contents of rats given rotenone: Rotenone in a dose of 2.5 mg/kg caused a significant increase in the concentrations of pro-inflammatory cytokines (TNF- α and IL-6) in the striatum, prefrontal cortex, and hippocampus of rats relative to controls (**Figures 9** and **10**). However, 100 mg/kg and 200 mg/kg SbS significantly reduced the concentrations of these pro-inflammatory cytokines in these brain regions of rats relative to rotenone (**Figures 9** and **10**).





Bars are mean \pm S.E.M. # p < 0.05 versus vehicle. * p < 0.05 versus rotenone (ANOVA followed by Bonferroni test) VEH=vehicle, SbS=Sorghum bicolor supplement, LD-CD=levodopa-carbidopa





Bars are mean±*S.E.M.* [#]*p*<0.05 versus vehicle. ^{*}*p*<0.05 versus rotenone (ANOVA followed by Bonferroni test) VEH=vehicle, SbS=Sorghum bicolor supplement, LD-CD=levodopa-carbidopa

Discussion

It is well established that intraperitoneal injection of rotenone induces parkinsonian-like motor deficits characterized by impaired locomotion, catalepsy and postural instability in rodents [3, 33-36]. Thus, it has served as an animal model widely used for the evaluation of novel agents with potential antiparkinsonian-like activity [3, 34-36]. In this study, the video analysis in the open field test confirmed that rotenone caused impairment of locomotor functions in rodents, evidenced by reduced distance traveled, number of lines crossed, navigational speed and increased freezing episodes/freezing time in the open field chamber. However, SbS at high doses attenuated rotenone-induced locomotor deficits in rats, suggesting potential benefits in ameliorating the motor symptoms associated with PD. Indeed, impairment of motor functions is a prominent feature of patients with PD and rats treated with rotenone [3, 4, 8, 35]. This impairment results in a reduction in voluntary movements and difficulty in initiating movement (akinesia) due to muscle rigidity [3]. The test for catalepsy is a common neurobehavioral feature that is widely measured in rodents that closely resemble akinesia in persons suffering from PD [37]. Catalepsy is defined as a state of motor dysfunction leading to difficulty in initiating voluntary movement characterized by the tendency of the limbs of the rats to remain in whatever position they are placed [37]. The results of the current study also support earlier investigation [37] which showed that intraperitoneal injection of rotenone-induced marked cataleptic behavior in rodents. Thus, the findings that SbS attenuated rotenone-induced catalepsy further suggest that it might offer beneficial effects in movement disorders pathognomonic of PD. It is worth noting that rotenone, a well-known neurotoxin, has been reported to cause neuropathological changes akin to PD via induction of oxidative stress due to disturbances in the mitochondrial electron transport system [3, 33, 34, 36]. Previous investigations have identified raised levels of oxidative stress and pro-inflammatory cytokines in the brain of animals after exposure to rotenone [3], and metabolic degradation of neuronal lipids and DNA [33, 38, 39]. Production of reactive oxygen and nitrergic species is well-known to be exacerbated by mitochondrial dysfunctions, microglia activation, neurodegeneration, and impaired antioxidant response mechanisms [3, 6, 38, 39]. Therefore, rotenone-induced depletion of endogenous antioxidant molecules (glutathione, catalase and superoxide dismutase) may contribute to its toxicity on dopaminergic neurons [38, 39]. It is known that a deficiency of striatal GSH has been reported in the literature and is believed to contribute to severe neuropathological derangements in PD [40]. In this study, SbS reduces MDA and nitrite concentrations and boosts antioxidant molecules in specific brain regions of rotenone-treated rats. Thus, these findings suggest that SbS exhibited an antioxidant defense protective effect against rotenone-induced motor deficits in rats.

Earlier investigations had also identified raised brain concentrations of pro-inflammatory cytokines in rodents exposed to rotenone [33, 36-38]. Indeed, several studies have shown that rotenone inhibited mitochondrial complex-I resulting in increased microglial activation, depletion of the antioxidant defense system and release of pro-inflammatory cytokines in various brain regions [33, 38]. Oxidative stress-mediated neuronal injury further releases pro-inflammatory cytokines, hence supporting the notion that oxidative stress and inflammation are co-conspirators in the demise of dopaminergic fibers [6, 38, 39]. It is interesting to state that SbS reduced the brain concentrations of pro-inflammatory cytokines induced by rotenone in rats. This finding is in agreement with earlier studies that showed that SbS mitigated neurological disorders in rats subjected to ischemic stroke via suppression of oxidative stress and pro-inflammatory cytokines [22]. Thus, the ability of SbS to reduce rotenone-induced Parkinsonian-like motor deficit in rats might also be related to its antioxidant and anti-inflammatory activities. Besides, it is also pertinent to mention that several studies have shown that SbS contains bioactive constituents, including apigenidin, apigenin, naringenin, and luteolin [19], with proven antioxidant, anti-inflammatory and neuroprotective activities [2, 16, 21, 41, 42].

Although previous studies have shown that SbS produces its most effective action between the doses of 50 mg/kg and 100 mg/kg, in this study, we intriguingly observed that SbS demonstrated a dose-dependent effect,

which suggests a unique therapeutic potential of SbS in the management of PD-like neuropathologies. As to whether SbS exerts superiority over LD-CD as a potential agent for motor impairment associated with rotenone, needs further investigation. However, in the context of safety and tolerability, it is important to mention that SbS is generally regarded as very safe for human consumption as it forms a very important part of our diets as a vegetable. Our previous research found that SbS is most effective at doses between 50 mg/kg and 100 mg/kg [22]. This study observed a dose-dependent effect of SbS of more than 100 mg/kg, indicating its unique stereoselective therapeutic potential against PD-like neuropathologies. However, more investigation is needed to determine if SbS is superior to LD-CD in addressing motor impairment associated with rotenone. It's worth noting that SbS is generally considered safe for human consumption and is a significant part of our diets as a vegetable. Interestingly, it was recently shown that SbS ameliorated liver and kidney impairment induced by aflatoxin-1 [43], a potent genotoxic hepatocarcinogen that causes up-regulation of alpha-synuclein, neuroinflammation, and degeneration of dopaminergic neurons [44]. However, the role(s) of phytochemicals in the anti-Parkinsonian-like effect of SbS requires further investigations.

Conclusion: This study suggests that SbS mitigated rotenone-induced parkinsonian-like motor dysfunctions by reducing oxidative stress and pro-inflammatory cytokines in rats indicating its therapeutic potential in movement-related disorders.

References

- 1. Pang SY-Y, Ho PW-L, Liu H-F, Leung C-T, Li L, Chang EES, Ramsden DB, Ho S-L (2019) The interplay of aging, genetics and environmental factors in the pathogenesis of Parkinson's disease. Translational Neuro-degeneration. 8: 23. doi: 10.1186/s40035-019-0165-9
- Yin R, Xue J, Tan Y, Fang C, Hu C, Yang Q, Mei X, Qi D (2021) The positive role and mechanism of herbal medicine in Parkinson's disease. Oxidative Medicine and Cellular Longevity. 9923331. ID: 1-23. doi: 10.1155/2021/ 9923331
- 3. Alam M, Schmidt WJ (2002) Rotenone destroys dopaminergic neurons and induces Parkinsonian symptoms in rats. Behavioural Brain Research. 136 (1): 317-324. doi: 10.1016/s0166-4328(02)00180-8
- 4. Olanow C, Schapira A (2013) Therapeutic prospects for Parkinson's disease. Annals of Neurology. 74 (3): 337-347. doi: 10.1002/ana.24011
- 5. Schapira AH, Jenner P (2011) Etiology and pathogenesis of Parkinson's disease. Movement Disorders. 26 (6): 1049-1055. doi: 10.1002/mds.23732
- 6. Taylor JM, Main BS, Crack PJ (2013) Neuroinflammation and oxidative stress: co-conspirators in the pathology of Parkinson's disease. Neurochemistry International. 62 (5): 803-819. doi: 10.1016/j.neuint.2012.12.016
- Sutachan JJ, Casas Z, Albarracin SL, Stab BR, Samudio I, Gonzalez J, Barreto G (2012) Cellular and molecular mechanisms of antioxidants in Parkinson's disease. Nutritional Neuroscience. 15 (3): 120-126. doi: 10.1179/ 1476830511Y.0000000033
- 8. Trist BG, Hare DJ, Double KL (2019) Oxidative stress in the aging substantia nigra and the etiology of Parkinson's disease. Aging Cell. 18 (6): e13031. doi: 10.1111/acel.13031
- 9. Nakabeppu Y, Tsuchimoto D, Yamaguchi H, Sakumi K (2007) Oxidative damage in nucleic acids and Parkinson's disease. Journal of Neuroscience Research. 85 (5): 919-934. doi: 10.1002/jnr.21191
- 10. Sarkar S, Raymick J, Imam S (2006) Neuroprotective and therapeutic strategies against Parkinson's disease: recent perspectives. International Journal of Molecular Sciences. 17 (6): 904. doi: 10.3390/ijms17060904
- Puspita L, Chung SY, Shim J-W (2007) Oxidative stress and cellular pathologies in Parkinson's disease. Molecular Brain. 10 (1): 53. doi: 10.1186/s13041-017-0340-9
- Bosco D, Fowler D, Zhang Q, Nieva J, Powers E, Wentworth P, Kelly J (2006) Elevated levels of oxidized cholesterol metabolites in Lewy body disease brains accelerate α-synuclein fibrilization. Nature Chemical Biology. 2 (5): 249-253. doi: 10.1038/nchembio782
- 13. Zeevalk GD, Razmpour R, Bernard LP (2008) Glutathione and Parkinson's disease: is this the elephant in the room? Biomedicine & Pharmacotherapy. 62 (4): 236-249. doi: 10.1016/j.biopha.2008.01.017
- 14. Hirsch E, Jenner P, Przedborski S (2008) Pathogenesis of Parkinson's disease. Movement Disorders. 28 (1): 24-30. doi: 10.1002/mds.25032

- 15. Schonhoff A, Williams G, Wallen Z, Standaert D, Harms A (2020) Innate and adaptive immune responses in Parkinson's disease. Progress in Brain Research. 252: 169-216. doi: 10.1016/bs.pbr.2019.10.006
- Sharma V, Bedi O, Gupta M, Deshmukh R (2022) A review: traditional herbs and remedies impacting pathogenesis of Parkinson's disease. Naunyn Schmiedeberg's Archives of Pharmacology. 395 (5): 495-513. doi: 10.1007/s00210-022-02223-5
- 17. Rai SN, Chaturvedi VK, Singh P, Singh BK, Singh MP (2020) *Mucuna pruriens* in Parkinson's and in some other diseases: recent advancement and future prospective. 3 Biotech. 10 (12): 522. doi: 10.1007/s13205-020-02532-7
- Rai SN, Singh P, Varshney R, Chaturvedi VK, Vamanu E, Singh MP, Singh BK (2021) Promising drug targets and associated therapeutic interventions in Parkinson's disease. Neural Regeneration Research. 16 (9): 1730-1739. doi: 10.4103/1673-5374.306066
- 19. Benson KF, Beaman JL, Ou B, Okubena A, Okubena O, Jensen GS (2013) West African *Sorghum bicolor* leaf sheaths have anti-inflammatory and immune-modulating properties in vitro. Journal of Medicinal Food. 16 (3): 230-238. doi: 10.1089/jmf.2012.0214
- 20. Umukoro S, Omogbiya IA, Eduviere AT (2013) Evaluation of the effect of jobelyn® on chemoconvulsantsinduced seizure in mice. Basic and Clinical Neuroscience. 4 (2): 125-129. PMID: 25337338.
- Oyinbo CA, Dare W, Avwioro O, Igbigbi P (2015) Neuroprotective effect of Jobelyn in the hippocampus of alcoholic rat is mediated in part by alterations in GFAP and NF 789 protein expressions. Advances in Biological Research. 9 (5): 305-317. doi: 10.5829/idosi.abr.2015.9.5.95109
- 22. Umukoro S, Oghwere EE, Ben-Azu B, Owoeye O, Ajayi AM, Omorogbe O, Okubena O (2019) Jobelyn® ameliorates neurological deficits in rats with ischemic stroke through inhibition of release of pro-inflammatory cytokines and NF-κB signaling pathway. Pathophysiology. 26 (1): 77-88. doi: 10.1016/j.pathophys.2018.10.00 2
- 23. John R, Abolaji AO, Adedara AO, Ajayi AM, Aderibigbe AO, Umukoro S (2022) Jobelyn® extends the life span and improves motor function in Drosophila melanogaster exposed to lipopolysaccharide via augmentation of antioxidant status. Metabolic Brain Disease. 37 (4): 1031-1040. doi: 10.1007/s11011-022-00919-4
- 24. Morais LH, Lima MM, Martynhak BJ, Santiago R, Takahashi TT, Ariza D, Barbiero JK, Andreatini R, Vital MA (2012) Characterization of motor, depressive-like and neurochemical alterations induced by a short-term rotenone administration. Pharmacological Reports. 64 (5):1081-1090. doi: 10.1016/s1734-1140(12)70905-2
- Arika WM, Kibiti CM, Njagi JM, Ngugi MP (2019) Effects of DCM leaf extract of Gnidia glauca (Fresen) on locomotor activity, anxiety, and exploration-like behaviors in high-fat diet-induced obese rats. Behavioural Neurology. 2019: 7359235. doi: 10.1155/2019/7359235
- 26. Costall B, Naylor R (1973) On catalepsy and catatonia and the predictability of the catalepsy test for neuroleptic activity. Psychopharmacologia. 34: 233-241. doi: 10.1007/BF00421964
- 27. Ohkawa H, Ohishi N, Yagi K (1979) Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Analytical Biochemistry. 95 (2): 351-358. doi: 10.1016/0003-2697(79)90738-3
- Green L, Tannenbaum S, Goldman P (1981) Nitrate synthesis in the germfree and conventional rat. Science. 212 (4490): 56-58. doi: 10.1126/science.6451927
- 29. Moron M, Depierre J, Mannervik B (1979) Levels of glutathione, glutathione reductase and glutathione S-transferase activities in rat lung and liver. Biochimica et Biophysica Acta. 582 (1): 67-78. doi: 10.1016/0304-4165(79)90289-7
- 30. Goth L (1991) A simple method for determination of serum catalase activity and revision of reference range. Clinica Chimica Acta. 196 (2-3): 143-151. doi: 10.1016/0009-8981(91)90067-m
- Misra H, Fridovich I (1972) The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. Journal of Biological Chemistry. 247 (10): 3170-3175. doi: 10.1016/S0021-9258(19) 45228-9
- 32. Lowry O, Rosebrough N, Farr A, Randall R (1951) Protein measurement with the folin phenol reagent. The Journal of Biological Chemistry. 193 (1): 265-275. PMID: 14907713.
- 33. Dhanalakshmi C, Janakiraman U, Manivasagam T, Justin Thenmozhi A, Essa M, Kalandar A, Guillemin G (2006) Vanillin attenuated behavioural impairments, neurochemical deficits, oxidative stress and apoptosis against rotenone induced rat model of Parkinson's disease. Neurochemical Research. 41 (8): 1899-1910. doi: 10.1007/s11064-016-1901-5
- Saeed A, Shakir L, Khan M, Ali A, Zaidi A (2017) Haloperidol induced Parkinson's disease mice model and motor-function modulation with Pyridine-3-carboxylic acid. Biomedical Research and Therapy. 4 (5): 1305-1317. doi: 10.15419/BMRAT.V4105.169
- 35. Cooper J, Spielbauer K, Senchuk M, Nadarajan S, Colaiácovo M, Van Raamsdonk J (2018) α-synuclein expression from a single copy transgene increases sensitivity to stress and accelerates neuronal loss in genetic models of Parkinson's disease. Experimental Neurology. 310: 58-69. doi: 10.1016/j.expneurol.2018.09.001

Mediterranean Journal of Pharmacy & Pharmaceutical Sciences



- www.medjpps.com
- 36. Waku I, Magalhaes M, Alves C, de Oliveira A (2021) Haloperidol-induced catalepsy as an animal model for parkinsonism: A systematic review of experimental studies. The European Journal of Neuroscience. 53 (11): 3743-3767. doi: 10.1111/ejn.15222
- 37. Alabi A, Ajayi A, Ben-Azu B, Bakre A, Umukoro S (2019) Methyl jasmonate abrogates rotenone-induced parkinsonian-like symptoms through inhibition of oxidative stress, release of pro-inflammatory cytokines, and down-regulation of immopositive cells of NF- κ B and α -synuclein expressions in mice. Neurotoxicology. 74: 172-183. doi: 10.1016/j.neuro.2019.07.003
- 38. Dawson TB, Dawson VL (2009) Molecular pathways of neurodegeneration in Parkinson's disease. Science. 302 (5646): 819-822. doi: 10.1126/science.1087753
- 39. Sherer T, Betarbet R, Testa CM, Seo BB, Richardson JR, Kim JH, Miller GW, Yagi T, Yagi AM, Greenamyre JT (2003) Mechanism of toxicity in rotenone models of Parkinson's disease. Journal of Neuroscience. 23 (34): 10756-10764. doi: 10.1523/JNEUROSCI.23-34-10756.2003
- 40. Pan P, Qiao L, Wen X (2016) Safranal prevents rotenone-induced oxidative stress and apoptosis in an in vitro model of Parkinson's disease through regulating Keap1/Nrf2 signaling pathway. Cellular and Molecular Biology. 62 (14): 11-17. PMID: 28145852.
- 41. Pearce R, Owen A, Daniel S, Jenner P, Marsden C (1997) Alterations in the distribution of glutathione in the substantia nigra in Parkinson's disease. Journal of Neural Transmission. 104 (6-7): 661-677. doi: 10.1007/ BF01291884
- 42. Afe TO, Alabi A, Ajayi AM, Ale AO, Fasesan OA, Ogunsemi OO (2024) Jobelyn® ameliorates anxiety response and oxido-inflammatory markers induced by tramadol use and discontinuation in rats. Mediterranean Journal of Pharmacy and Pharmaceutical Sciences. 4 (1): 93-110. doi: 10.5281/zenodo.10728692
- 43. Owumi S, Kazeem A, Wu B, Ishokare L, Arunsi U, Oyelere A (2022) Apigeninidin-rich Sorghum bicolor (L. Moench) extracts suppress A549 cells proliferation and ameliorate toxicity of aflatoxin B1-mediated liver and kidney derangement in rats. Scientific Reports. 12 (1): 7438. doi: 10.1038/s41598-022-10926-1
- 44. Wang W, Wang Y, Wagner K, Lee R, Hwang S, Morisseau C, Wulff H, Hammock B (2023) Aflatoxin B₁ increases soluble epoxide hydrolase in the brain and induces neuroinflammation and dopaminergic neurotoxicity. International Journal of Molecular Sciences. 24 (12): 9938. doi: 10.3390/ijms24129938

Ethical issues: Including plagiarism, informed consent, data fabrication or falsification, and double publication or submission were completely observed by the authors.

Data availability statement: The raw data that support the findings of this article are available from the corresponding author upon reasonable request.

Author declarations: The authors confirm that all relevant ethical guidelines have been followed and any necessary IRB and/or ethics committee approvals have been obtained.

Acknowledgments: The authors wish to thank the technical staff of the Department of Pharmacology & Therapeutics, University of Ibadan, Nigeria for their assistance during the period of the experiments.

Authors' contributions: PAA, OSA, OO & SU conceived and designed the study. PAA, OSA & AMA collected the data. OSA, AMA & PAA conducted experiments. SU, OSA, and BB analyzed and interpreted the data. SU & BB wrote the manuscript. All authors read and approved the final version of the manuscript. The authors declare that all data were generated in-house and that no paper mill was used.

Conflict of interest: The authors declare the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.